

## Notes

FUJIANMYCINS A AND B,  
NEW BENZ[a]ANTHRAQUINONE  
ANTIBIOTICS FROM A  
*STREPTOMYCES* SPECIES

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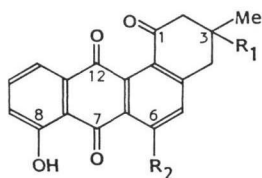
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The small group of antibiotics from *Streptomyces* species which are derivatives of the angularly-fused benz[a]anthraquinone structure includes tetrangomycin (**1**)<sup>1,2</sup>, its aromatization product tetrangulol<sup>1,2</sup>, and its 6-hydroxy-analogue rabelomycin (**2**)<sup>3</sup>. From an unidentified *Streptomyces* species, obtained from a soil sample collected in Fujian Province during the course of a screening program conducted by the Institute of Antibiotics of the Chinese Academy of Medical Sciences, we have obtained two new members of this benz[a]anthraquinone group. We describe here the production, isolation and structural analysis of these compounds, named fujianmycins A (**4**) and B (**5**).

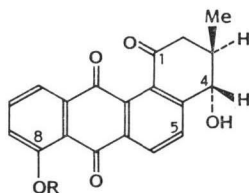
The inoculum of the *Streptomyces* sp. (IA-CAS isolate No. 114) was prepared in a medium containing beef extract 0.5%, mycological peptone 0.5%, yeast extract 0.5%, enzymatic digest of casein 0.3%, sodium chloride 0.15%, glucose 2.0% in distilled water adjusted to pH 7.5. Antibiotics were produced by growth at 28°C in

baffled conical flasks (250 ml) containing a medium (100 ml) consisting of mycological peptone 0.4%, yeast extract 0.1%, Bacto-soytone 0.5%, glucose 2.5%, calcium carbonate 0.2% in distilled water. After fermentation for 72 hours, the broth (pH 7.8) was adjusted to pH 6.5 with hydrochloric acid (8 N) and filtered. The filtrate (7.8 liters) was extracted ( $\times 3$ ) with equal volumes of EtOAc, and the extracts were dried and evaporated. The resulting tar (929 mg) in  $\text{CH}_2\text{Cl}_2$  (100 ml) was washed successively with cold  $\text{NaHCO}_3$  (2%,  $2 \times 100$  ml) and water ( $2 \times 50$  ml), dried, and re-evaporated. The residue (116 mg) was chromatographed on Sephadex LH-20 in MeOH -  $\text{CHCl}_3$  (1:99) to give a yellow fraction (32 mg) containing fujianmycins A and B. Preparative TLC on silica gel in toluene - EtOAc (3:7) gave fujianmycin A (10 mg) and fujianmycin B (9.8 mg).

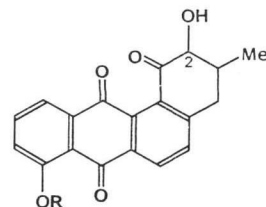
Fujianmycin A formed yellow needles from  $\text{CH}_2\text{Cl}_2$  - hexane (1:1), mp 182~185°C. MS  $m/z$  322.0817 (M,  $\text{C}_{10}\text{H}_{14}\text{O}_5$  requires 322.0841), 304(M-H<sub>2</sub>O), 294(M-CO), 280.0349 (M-CH<sub>2</sub>-CHMe,  $\text{C}_{10}\text{H}_8\text{O}_5$  requires 280.0372), 264.0410 (M-CHOHCHMe,  $\text{C}_{10}\text{H}_8\text{O}_4$  requires 264.0423), 251.0331 (M-C<sub>4</sub>H<sub>7</sub>O,  $\text{C}_{10}\text{H}_7\text{O}_4$  requires 251.0344); UV  $\lambda_{\text{max}}^{\text{MeOH}}$  nm 273 and 402, altered to 267, 326 and 490 on addition of NaOH; IR  $\lambda_{\text{max}}$  ( $\text{CH}_2\text{Cl}_2$  -  $\text{CCl}_4$ , 3:7)  $\text{cm}^{-1}$  1640 (H-bonded quinone C=O), 1678 (quinone C=O), 1710 (ketonic C=O), 2500~3300 (H-bonded OH);  $[\alpha]_{\text{D}}^{20} +17^\circ$  ( $c \sim 0.08$ ,  $\text{CHCl}_3$ );  $^1\text{H}$  NMR (200 MHz,  $\text{CDCl}_3$ )  $\delta$  1.26 (3H, d,  $\text{CHCH}_3$ ), 2.35 (1H, m, 3-H), 2.58 (1H, dd,  $J_{2\text{ax},2\text{eq}}=16.4$  Hz,  $J_{2\text{ax},3}=10.3$  Hz, 2-H<sub>ax</sub>), 3.14 (1H, dd,  $J_{2\text{eq},2\text{ax}}=16.4$  Hz,  $J_{2\text{eq},3}=$



- 1**  $\text{R}_1 = \text{OH}$ ,  $\text{R}_2 = \text{H}$   
**2**  $\text{R}_1 = \text{R}_2 = \text{OH}$   
**3**  $\text{R}_1 = \text{R}_2 = \text{H}$



- 4**  $\text{R} = \text{H}$   
**5**  $\text{R} = \text{Me}$



- 6**  $\text{R} = \text{H}$  or  $\text{Me}$

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6.1 Hz, 2-H<sub>eq</sub>), 4.51 (1H, d,  $J_{4ax,3} = 9.3$  Hz, 4-H<sub>ax</sub>), 7.29 (1H, dd,  $J_s$  not readable due to CHCl<sub>3</sub>, 9-H), 7.68 (2H, m, 10-H and 11-H), 8.09 (1H, d with slight broadening,  $J_{5,0} = 8.3$  Hz, 5-H), 8.41 (1H, d,  $J_{6,5} = 8.3$  Hz, 6-H), 12.30 (1H, s, exchanged with D<sub>2</sub>O, H-bonded OH).

Fujianmycin B crystallized as yellow needles from CH<sub>2</sub>Cl<sub>2</sub> - hexane (1:1), mp 233~235°C.

Anal Calcd for C<sub>20</sub>H<sub>10</sub>O<sub>5</sub>: C 71.4, H 4.8.

Found: C 71.7, H 5.6.

MS  $m/z$  336.1005 (M, C<sub>20</sub>H<sub>10</sub>O<sub>5</sub> requires 336.0998), 318 (M-H<sub>2</sub>O), 308 (M-CO), 294.0528 (M-CH<sub>2</sub>-CHMe, C<sub>17</sub>H<sub>10</sub>O<sub>5</sub> requires 294.0528), 278.0562 (M-CHOHCHMe, C<sub>17</sub>H<sub>10</sub>O<sub>4</sub> requires 278.0579), 265.0505 (M-C<sub>4</sub>H<sub>7</sub>O, C<sub>16</sub>H<sub>9</sub>O<sub>4</sub> requires 265.0501); UV  $\lambda_{max}^{MeOH}$  nm ( $\epsilon$ ) 273 (22,850) and 382 (3,360), unaltered on addition of NaOH; IR  $\lambda_{max}$  (CH<sub>2</sub>-Cl<sub>2</sub> - CCl<sub>4</sub>, 1:1) cm<sup>-1</sup> 1675 (quinone C=O), 1705 (ketonic C=O), 3600 (OH);  $[\alpha]_D^{25} + 50.0^\circ$  ( $c$  0.176, CHCl<sub>3</sub>); <sup>1</sup>H NMR (200 MHz, CDCl<sub>3</sub>)  $\delta$  1.26 (3H, d,  $J = 6.6$  Hz, CHCH<sub>3</sub>), 2.35 (1H, m, 3-H), 2.56 (1H, dd,  $J_{2ax,2eq} = 16.4$  Hz,  $J_{2ax,3} = 10.7$  Hz, 2-H<sub>ax</sub>), 3.09 (1H, dd,  $J_{2eq,2ax} = 16.4$  Hz,  $J_{2eq,3} = 5.6$  Hz, 2-H<sub>eq</sub>), 4.05 (3H, s, OCH<sub>3</sub>), 4.50 (1H, d,  $J_{4ax,3} = 9.2$  Hz, 4-H<sub>ax</sub>), 7.31 (1H, dd,  $J_{9,10} = 7.9$  Hz,  $J_{9,11} = 1.5$  Hz, 9-H), 7.71 (1H, t,  $J_{10,9} \approx J_{10,11} \approx 7.8$  Hz, 10-H), 7.77 (1H, dd,  $J_{11,10} = 7.7$  Hz,  $J_{11,9} = 1.6$  Hz, 11-H), 8.00 (1H, d with slight broadening,  $J_{5,6} = 8.2$  Hz, 5-H), 8.34 (1H, d,  $J_{6,5} = 8.2$  Hz, 6-H).

These spectral data indicate that the neutral fujianmycin B is the methyl ether of the phenolic fujianmycin A. In confirmation, methylation of fujianmycin A (MeI-Ag<sub>2</sub>O, CHCl<sub>3</sub>, room temp, 12 hours) gave an *O*-methyl derivative identical in R<sub>f</sub> value on TLC, mass and <sup>1</sup>H NMR spectra with fujianmycin B, together with a minor *O,O*-dimethyl derivative (MS  $m/z$  350 (M, C<sub>22</sub>H<sub>12</sub>O<sub>5</sub>)) in which an alcoholic hydroxyl function has also been methylated.

The chromophore of both fujianmycins in neutral solution closely resembles that of the 3,4-dihydro-8-hydroxy-benz[*a*]anthracene-1,7,12-(2*H*)-trione tetrangomycin (**1**)<sup>1,2)</sup>, which is isomeric with fujianmycin A. (The chromophores of the phenols fujianmycin A and tetrangomycin in alkaline solution do not correspond, since tetrangomycin undergoes rapid base-catalysed dehydration and aromatization to tetrangulol under these conditions<sup>1,2)</sup>, in contrast to the present compounds which are stable). As expected, fujianmycin A shows IR absorption

maxima corresponding to the ketonic carbonyl and the free and H-bonded quinone carbonyl groups of tetrangomycin (**1**). Furthermore, <sup>1</sup>H NMR spectra of both fujianmycins show the two- and three-proton vicinal aromatic spin systems of tetrangomycin (**1**), while fujianmycin A shows a strongly hydrogen-bonded phenolic group at  $\delta$  12.30 which is methylated in fujianmycin B. The remaining eight protons of the fujianmycins are contained in the system -CH<sub>2</sub>CHMeCHOH-, for which assignments and spin coupling constants (fully confirmed by double resonance experiments) are presented in the data above. This segment, linked to the carbonyl group located at C-1 from electronic spectra, must form the angularly-fused fourth ring of the fujianmycins. Two orientations of the segment are possible, leading to the 4-hydroxyl structures **4** and **5** or to structures of the 2-hydroxyl type **6**. The observation in <sup>1</sup>H NMR spectra of fujianmycin B of an 0.8 Hz coupling between the methine proton of the secondary alcohol and the aromatic proton at C-5 confirms the position of the carbonyl group and established the structures **4** and **5** for the fujianmycins. This coupling, which is clearly defined in resolution-enhanced spectra, is removed only upon irradiation of the benzylic methine proton.

The *trans* relative configuration of the methyl and hydroxyl substituents of the fujianmycins (**4**) and (**5**) follows from the magnitude of the proton coupling constants<sup>4)</sup> in their hydroaromatic rings, particularly the two large (9~11 Hz) couplings between the three *quasi*-axial protons at C-2, C-3 and C-4. The two antibiotics must have the same absolute configuration, since they have positive D-line rotations and effectively the same chromophore.

Fujianmycin A (**4**) showed antibacterial activity in disc assays against *Bacillus subtilis* on seeded agar plates at 50  $\mu$ g/ml. In contrast, its methyl derivative, fujianmycin B (**5**) was active only at much higher concentrations, indicating the importance of the phenolic function for antimicrobial activity.

Another benz[*a*]anthraquinone from *Streptomyces* species, ochromycinone, was suggested to have either the 8-hydroxyl structure **3** or the corresponding 11-hydroxyl structure, the former being preferred on biogenetic grounds<sup>5)</sup>. Disregarding questions of absolute configuration at C-3, structure **3** represents a deoxy-derivative of both tetrangomycin (**1**) and fujianmycin A (**4**),

and this 8-hydroxyl formulation for ochromycinone is confirmed by the close correspondence of the electronic spectra of these three phenols:  $\lambda_{\max}^{\text{MeOH}}$  nm **1** 267, 400; **3** 265, 405; **4** 273, 402.

The anthraquinones fujianmycins A (**4**) and B (**5**), ochromycinone (**3**), tetrangomycin (**1**), rabelomycin (**2**) and tetrangulol are clearly related biosynthetically to the naphthoquinones of the aquayamycin<sup>6)</sup>, yonomycin<sup>7)</sup>, P-1894 B<sup>8,9)</sup>, vineomycin<sup>9,10)</sup>, and sakyomicin<sup>11,12)</sup> group, in which the [a]-ring junction bond of the 3,4-dihydrobenz[a]anthracene-1,7,12(2*H*)-trione system has been oxidised to a 4a,12b-diol.

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